



DEPTH FILTRATION FOR VALUABLE LIQUIDS. SINCE 1938.

NEW APPROACH FOR SINGLE-USE CLARIFICATION OF BIOLOGICAL SOLUTIONS LEVITRONIX CONFERENCE JUNE 2022

Bettina Ledergerber, Cambridge, 02.04.2022



Bettina Ledergerber

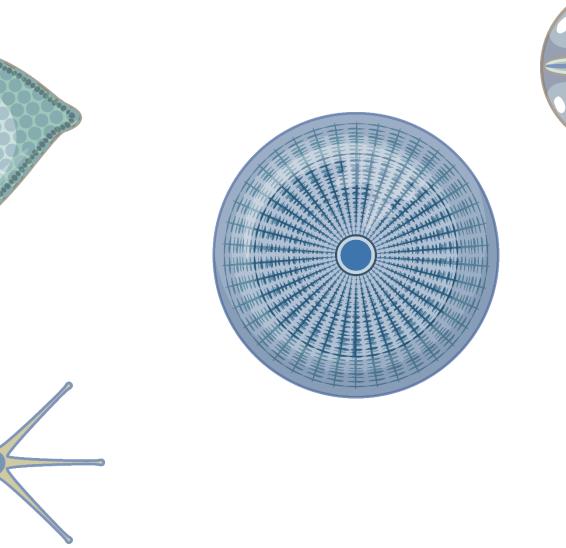
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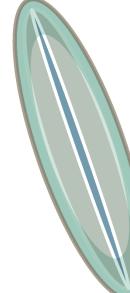




ONCE UPON A TIME...





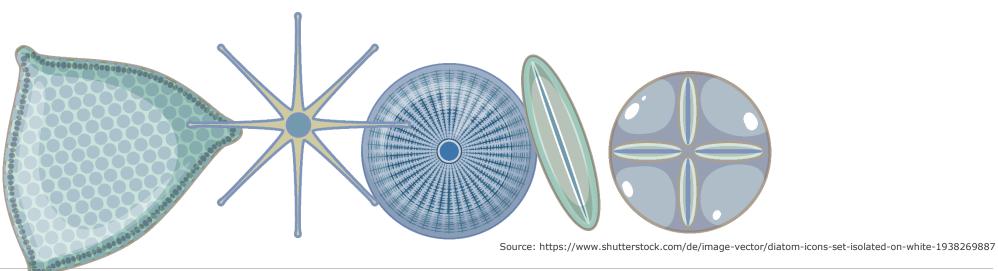


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ONCE UPON A TIME...





AGENDA



- Trends & Technologies in biomanufacturing
- Sheet vs. Depth vs. Alluvial filtration

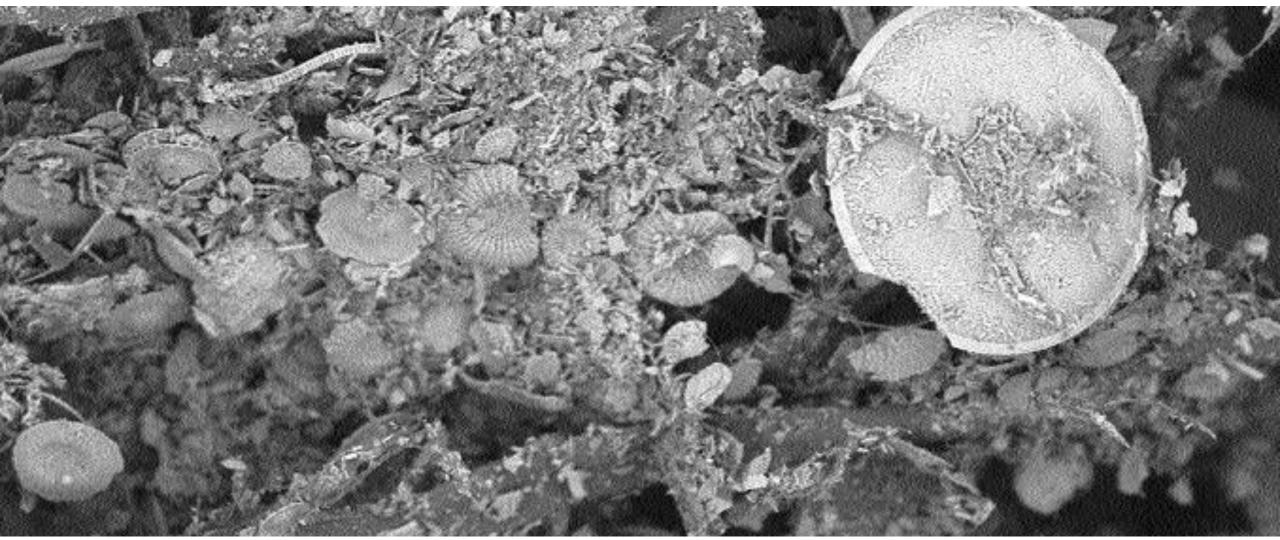
Case Studies

- 1) Mammalian Cell removal
- 2) Plasmid DNA clarification
- 3) Depth Filtration vs. Alluvial filtration vs. pH adjustment
- 4) Influence of different filter sheet types
- 5) Scale up



BASICS OF DEPTH FILTRATION

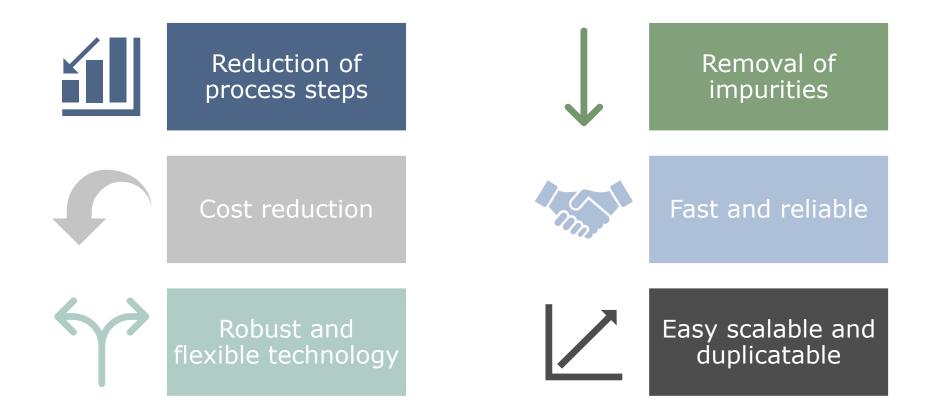




Source: FILTROX AG, 2008

TRENDS

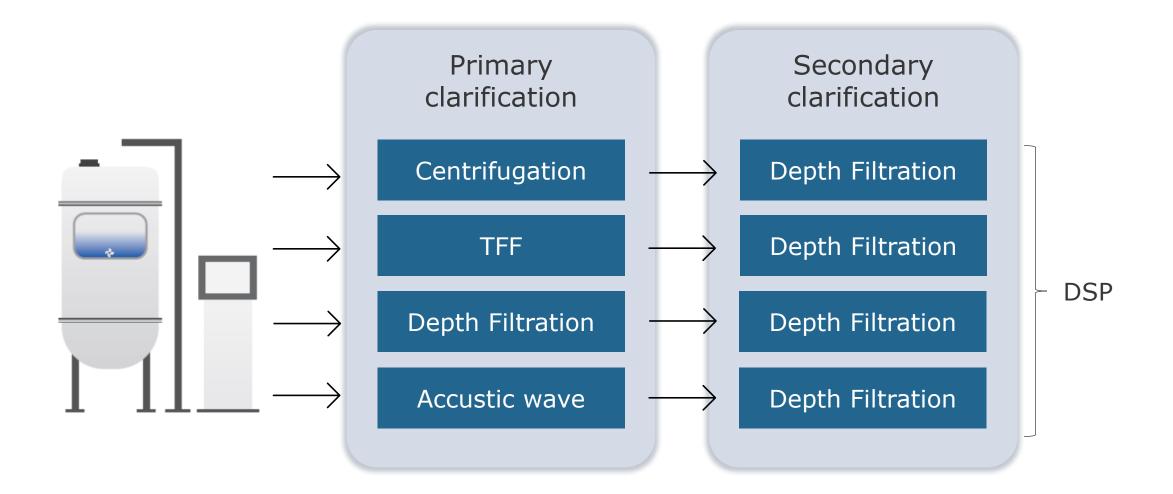
Development and challenges in Biomanufacturing





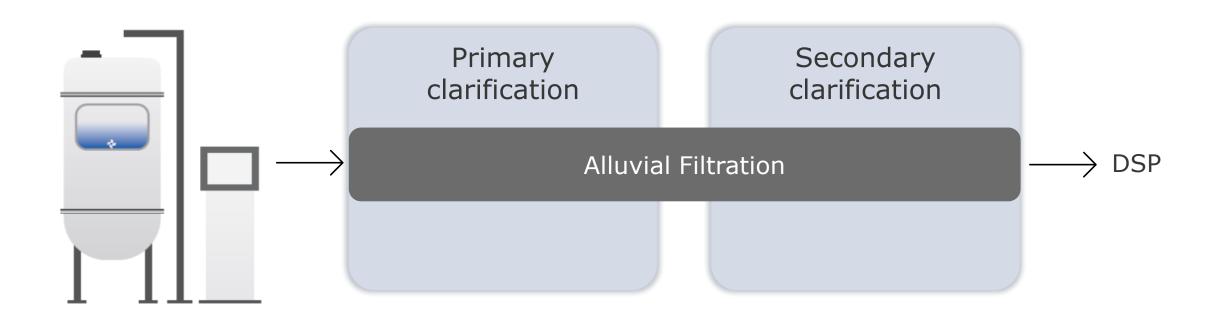
TECHNOLOGY COMPARISON





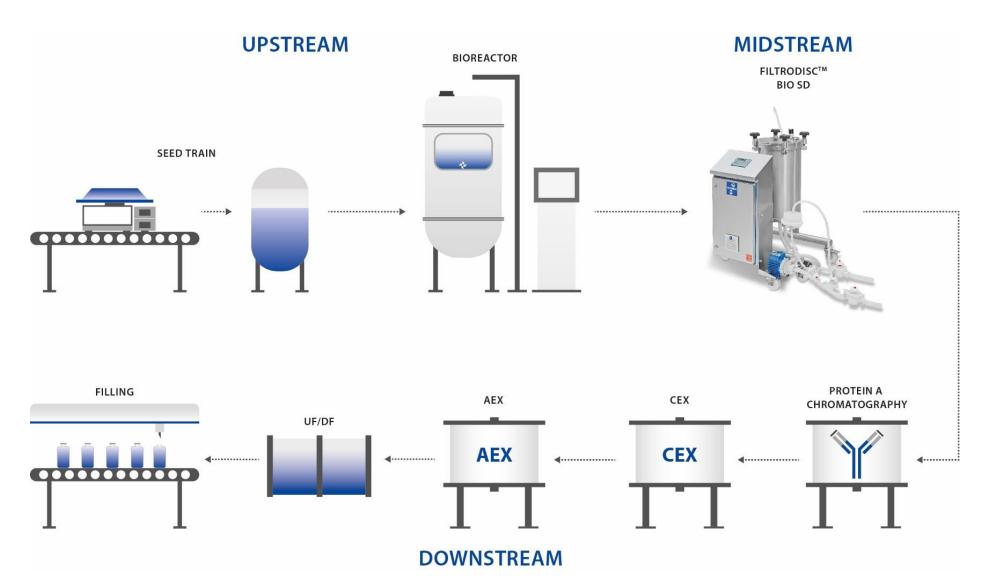
TECHNOLOGY COMPARISON





TECHNOLOGY COMPARISON

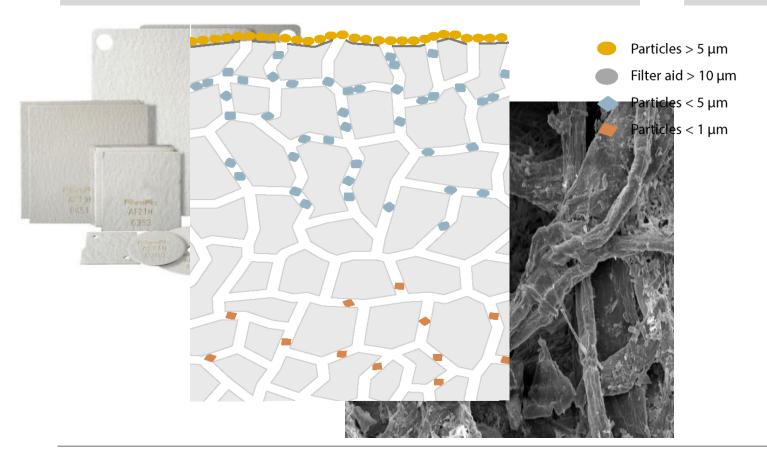




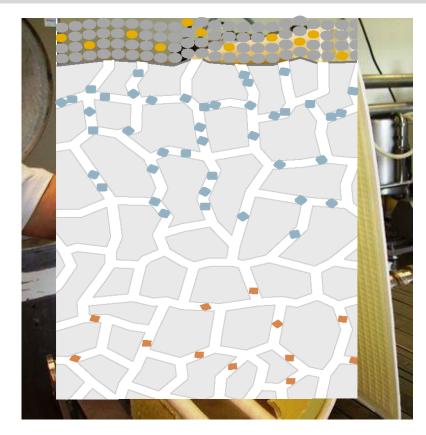
DEPTH VS. ALLUVIAL FILTRATION



Depth Filtration

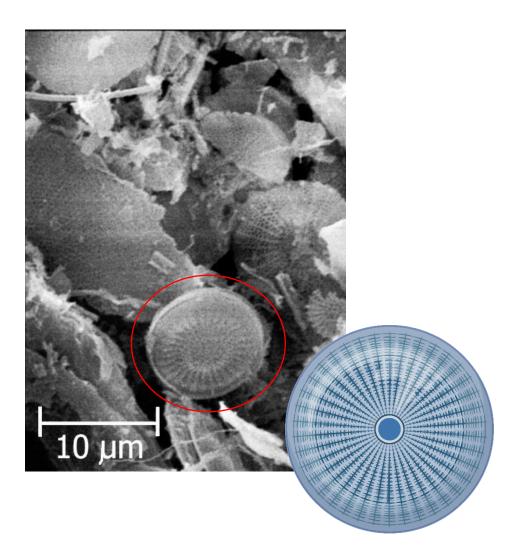


Alluvial Filtration



- Large Capacity for high particle load
- An established technology in pharmaceutical filtration since decades (e. g. plasma fractionation)
- Long term experience (over 60 years in beer filtration)
- Filtration through 3D Network
- Economical
- → Constantly renewed filter surface



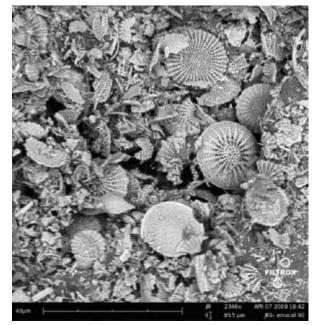


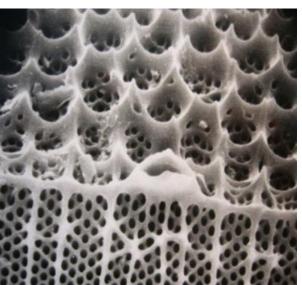


Diatomaceous earth = DE = Kieselguhr

- E. g. Celpure[®] (an imerys brand) \rightarrow pharma grade
- Inorganic filter aid
- Mineralized organisms (skeleton of diatoms)

Grade	Permebaility [mDarcy]	Solids removed* [µm]
C65	40 - 80	0.3 - 0.45
C100	70 - 140	0.3 - 0.45
C300	150 - 300	0.45 - 0.6
C1000	750 – 1250	> 1.0

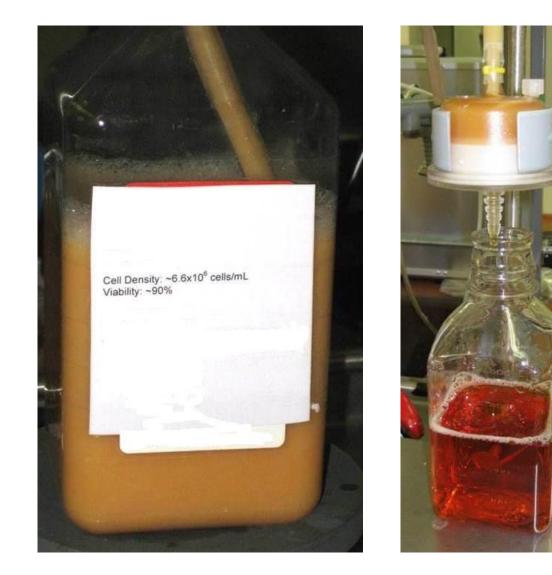




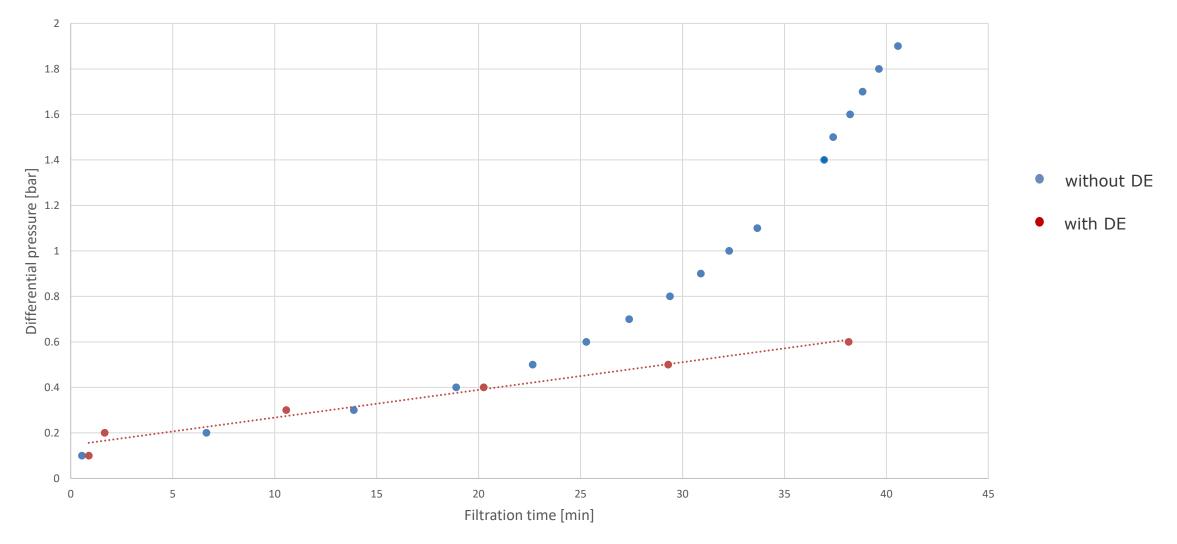
FILTROX

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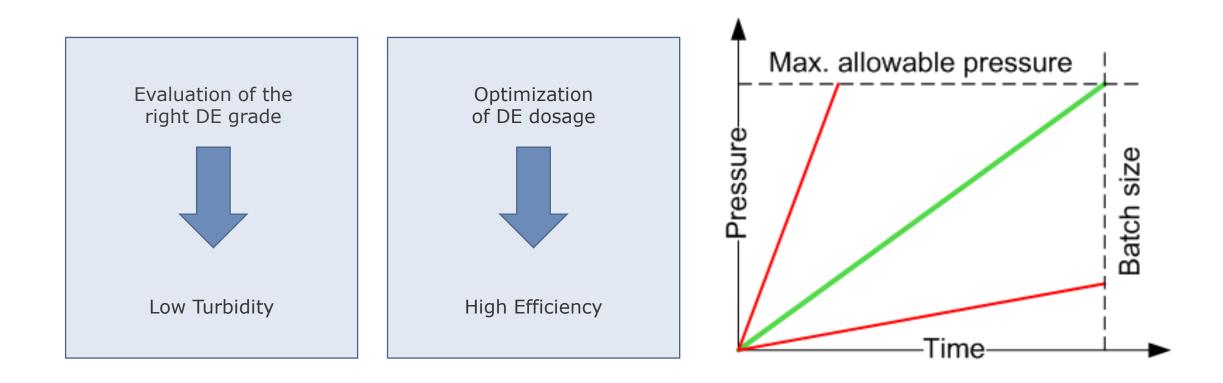






FILTRODISC[™] BIO SD





FILTRODISC[™] BIO SD





AGENDA



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- Sheet vs. Depth vs. Alluvial filtration

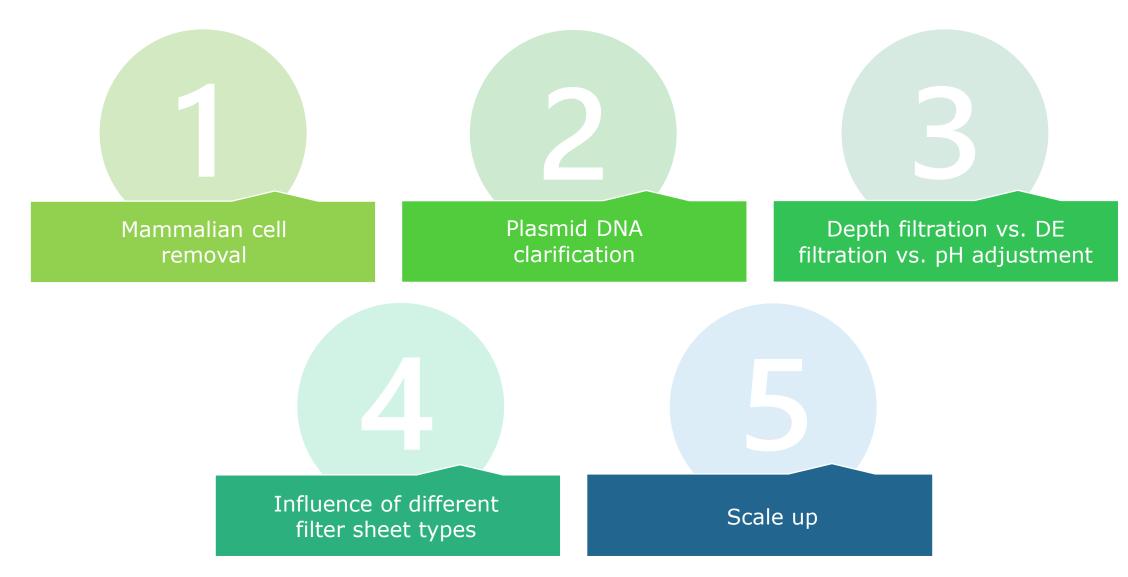
Case Studies

- 1) Mammalian Cell removal
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- 3) Depth Filtration vs. Alluvial filtration vs. pH adjustment
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- 5) Scale up



CASE STUDIES



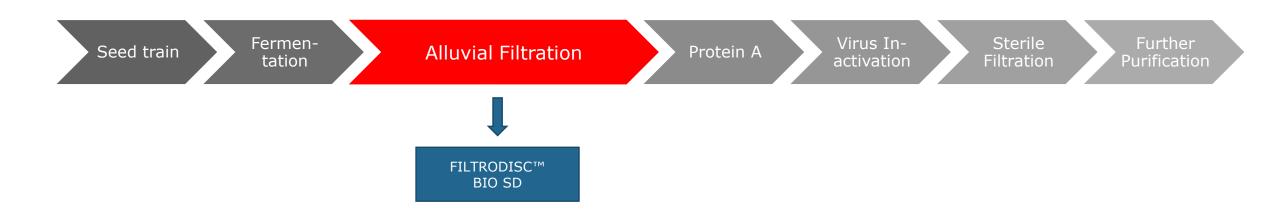


CASE STUDY: MAMMALIAN CELL REMOVAL



Background:

Typical clarification process for mammalian cells involves primary and secondary clarification step \rightarrow Process step reduction with FILTRODISCTM BIO SD system and alluvial filtration technology



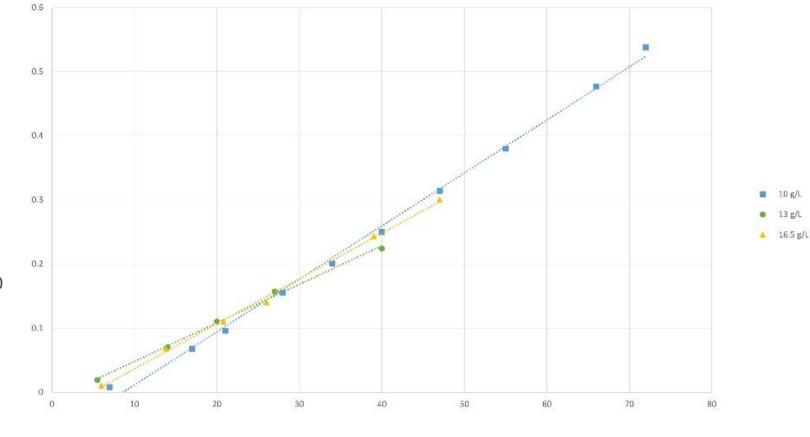
CASE STUDY: MAMMALIAN CELL REMOVAL

CHO cell culture producing IgG

- 2 days fed-batch cultivation
- 80 99 % viability
- $5 6 \times 10^6$ cells
- Harvest strategy:
 - FILTRODISC[™] BIO SD 2"
 - PURAFIX[®] CH 09P

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- Comparison of Celpure[®] C100 and C300
- Comparison of different Celpure[®]
 amounts
- Filtration flux = $400 500 \text{ L/m}^2 \times \text{h}$



Filtration time (min)

Figure: Pressure profiles as a function of throughput for different dosage of Celpure[®] 300



Pressure (bar)



CASE STUDY: MAMMALIAN CELL REMOVAL

CHO cell culture producing IgG

→ Scale up from FILTRODISC[™] BIO SD 2" to 5"

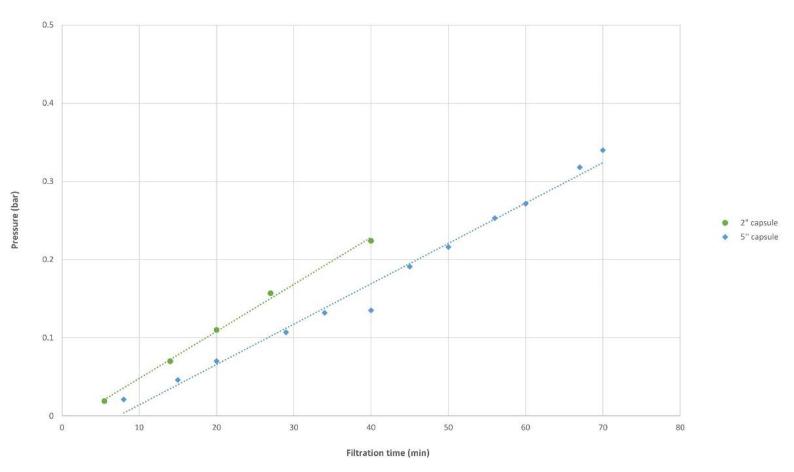


Figure: Pressure profiles (inlet) as a function of process time (throughput) for two different filtration FILTRODISC[™] BIO SD 2" and 5" capsules and 13.5 g/L DE dose

CASE STUDY: MAMMALIAN CELL REMOVAL

CHO cell culture producing IgG

 \rightarrow Recovery Yield

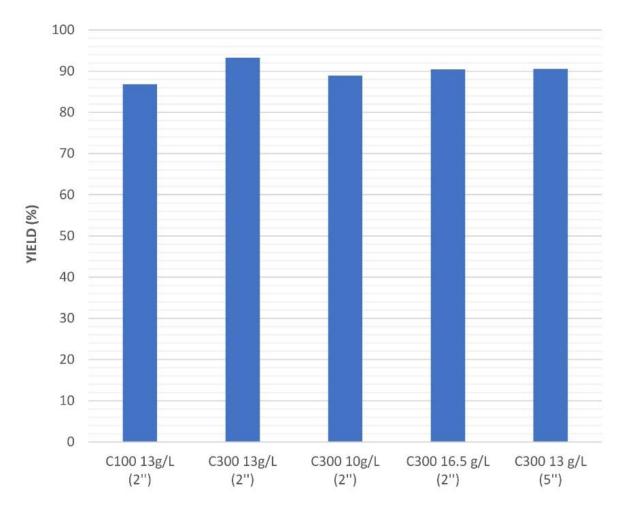


Figure: IgG recoveries in different filtration experiments



CASE STUDY: MAMMALIAN CELL REMOVAL

CHO cell culture producing IgG

→Results

- Productive approach
 - High flux (450 L/m²×h)
 - High throughput (400 L/m² as standard)
- Consistent clarification performance
 - Filtration fluxes
 - Throughputs
 - Reduction of turbidity
- Simple scale up
- Superior performance in terms of capacity & productivity (compared to alternative technologies)
- Cost efficiency \rightarrow important for processes that are associated with "acceptance criteria" in terms of manufacturing costs

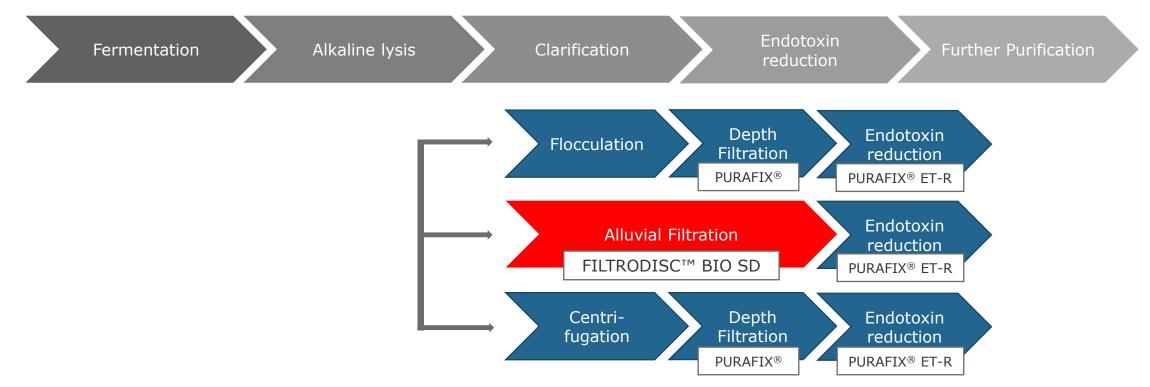
CASE STUDY: PLASMID DNA CLARIFICATION



Background:

Plasmid DNA is used in production of viral vectors.

→ Increasing demand for plasmid DNA leads to need for larger manufacturing capacities (due to increased clinical success and growing number of late-phase clinical studies)

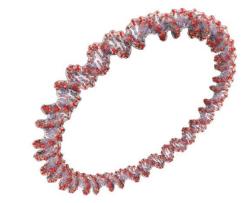


CASE STUDY: PLASMID DNA CLARIFICATION



→Advantages

- Reduction of process steps (by elimination of centrifugation step)
- Process time reduction
- Cost effective reduction of impurities
- Flexible in terms of space for filter cake, retention rate, connectors etc.

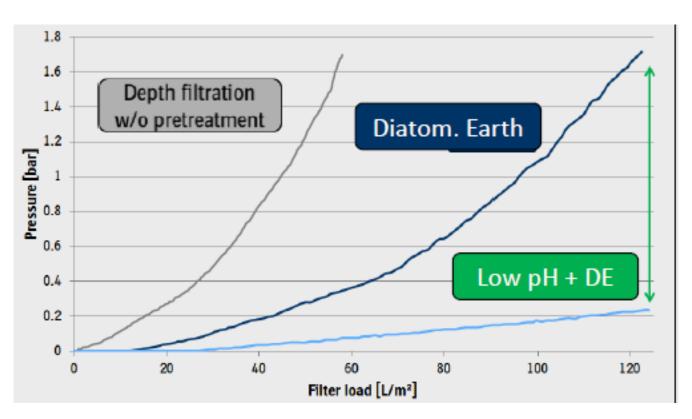




CASE STUDY: DEPTH FILTRATION VS. DE FILTRATION VS. ACID PRECIPITATION

Acid treatment / low pH:

- Leads to bigger particles
- Easier to filter
- Influence on target molecule?
- Afterwards pH needs to be adjusted again



Data published by Dr. Markus Brakel, Boehringer Ingelheim, BioProduction 2017 Dublin



CASE STUDY: INFLUENCE OF DIFFERENT DEPTH FILTER SHEET TYPES



Background:

The use of filter sheets with different characteristics (retention rate) have an influence on the turbidity

Conditions:

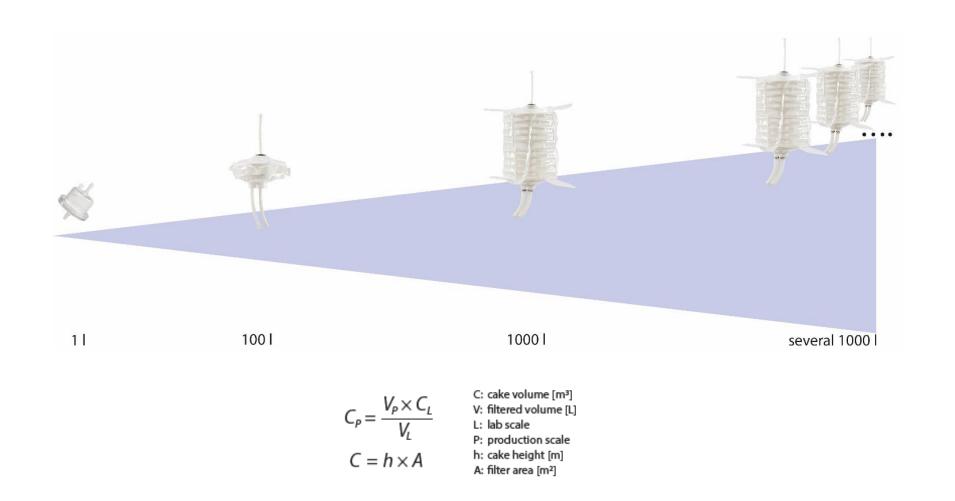
- 6×10^6 cells / mL
- Viability: 90 %
- Turbidity: 295 NTU
- DE grade: Celpure[®] C1000
- DE dosage: 25 g/L

	Trial 1	Trial 2
Filter sheet grade	PURAFIX [®] CH 9P	PURAFIX [®] CH ST 130P
Renteion rate [µm]	30 - 10	0.6 - 0.4
Average flux [L/m ² ×h]	648	420
Turbidity after filtration [NTU]	3.8	1.5



CASE STUDY: SCALE UP





CASE STUDY: SCALE UP



Background:

Conditions:



- Laboratory scale filtration of a CHO cell culture
- FILTRODISC[™] BIO SD 2" capsule (21 cm²)
 - Filtered volume = 380 mL
 - Final ΔP = 2.3 bar
 - Filter aid type = Celpure[®] C300
 - Filter aid dosage = 50 g/L
 - Cake height = 3.0 cm
 - Batch size = 100 L

Scale up:

$$A_{prod.}[m^{2}] = \frac{V_{prod.}[L] \times A_{test}[m^{2}]}{V_{test}[L]} = \frac{100 \ L \times 0.0021 \ m^{2}}{0.38 \ L} = 0.55 = 0.6 \ m^{2}$$
$$c_{prod.}[m^{3}] = \frac{V_{prod.}[L] \times c_{test}[m^{3}]}{V_{test}[L]} = \frac{100 \ L \times 0.000063 \ m^{3}}{0.38 \ L}$$
$$= 0.0166 \ m^{3} = 16.6 \ L$$

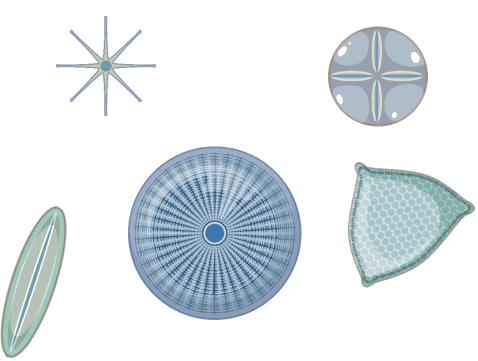
→ FILTRODISCTM BIO SD 12" double module

- 6 lenses per module (overall 12) = 1.32 m²
- Max holding volume = 17.8 L

TAKE-HOME MESSAGE

Advantages of Alluvial Filtration technology

- Well know and accepted technology (e. g. plasma fractionation)
- Removes cells and impurities in one step
- No need for pH changes or flocculants
- Reduce process time
- Different DE types available









DEPTH FILTRATION FOR VALUABLE LIQUIDS. SINCE 1938.

Let us solve your filtration task!

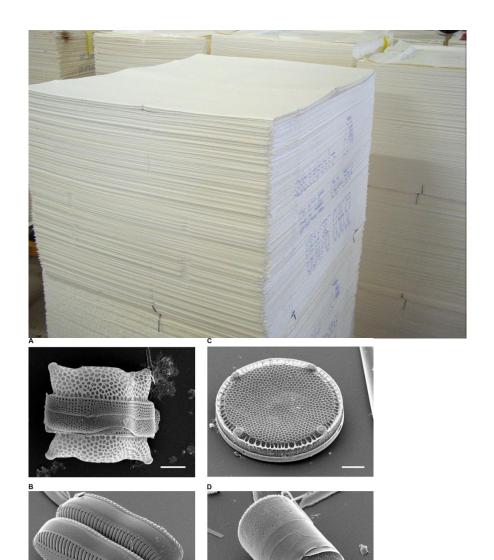
APPENDIX

- Filter sheet composition
- Filter sheet production process
- Validation
- FILTRODISC[™] BIO SD series
- FILTRODISC[™] BIO SD: Filtration principle
- Laboratory scale set-up
- Powder Transfer
- Case Study: Viral Vector Clarification



FILTER SHEET COMPOSITION

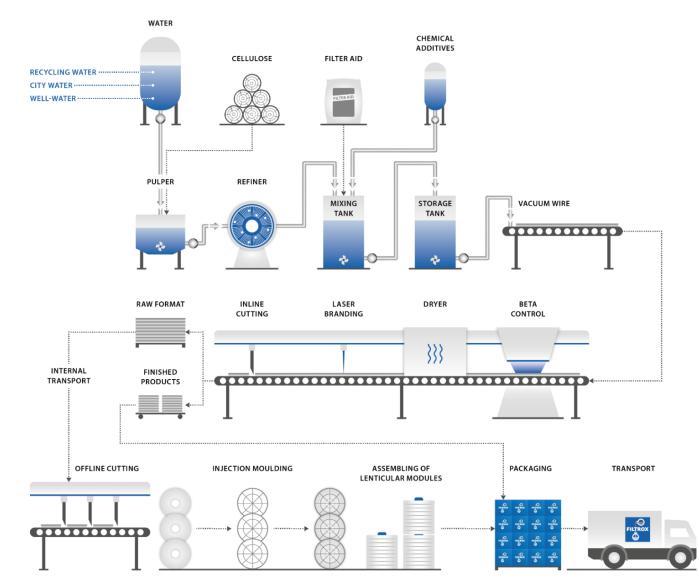
- Cellulose fibers
 - Ground cellulose fibers form a 3D matrix
- Filter aid (e. g. Kieselguhr, Perlite)
 - Enlarge the inner surface
 - Improve the filtration effects
- Wet strength agent
 - Increase of wet tensile strength
 - Positive charge for the adsorption of negatively charged suspended solids





FILTER SHEET PRODUCTION PROCESS





PRIMARY PRODUCTION



Pulper Wet sieve section

Laser marking

Drying Oven

SECONDARY PRODUCTION





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PRODUCTION PROCESS



laser marking)

Unfiltrat (coarse surface)





VALIDATION

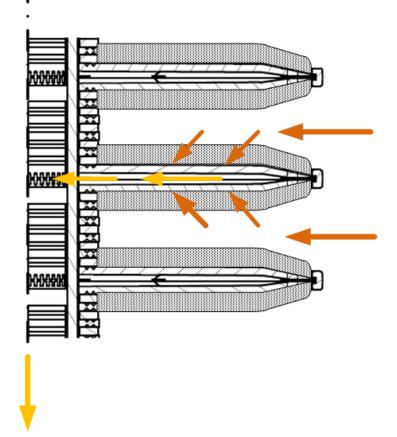


Material of Construction

Supporting filter sheets:	PURAFIX [®] low ion and low pyrogen containing filter sheets
Filter aid:	Pharma grade Diatomaceous earth Celpure [®] ; DE is already part of some sheet types
Bag material:	Double layer PE material, already used for other single-use applications (e. g. Bioreactors)
Tube:	APSH - Platinum Cured Silicone
Other plastic parts:	USP class VI material (e. g. core body and drainage nets of the modules)
Powder transfer bags:	LDPE with Permanent Anti-Static Additive

FILTRODISC[™] BIO SD: FILTRATION PRINCIPLE



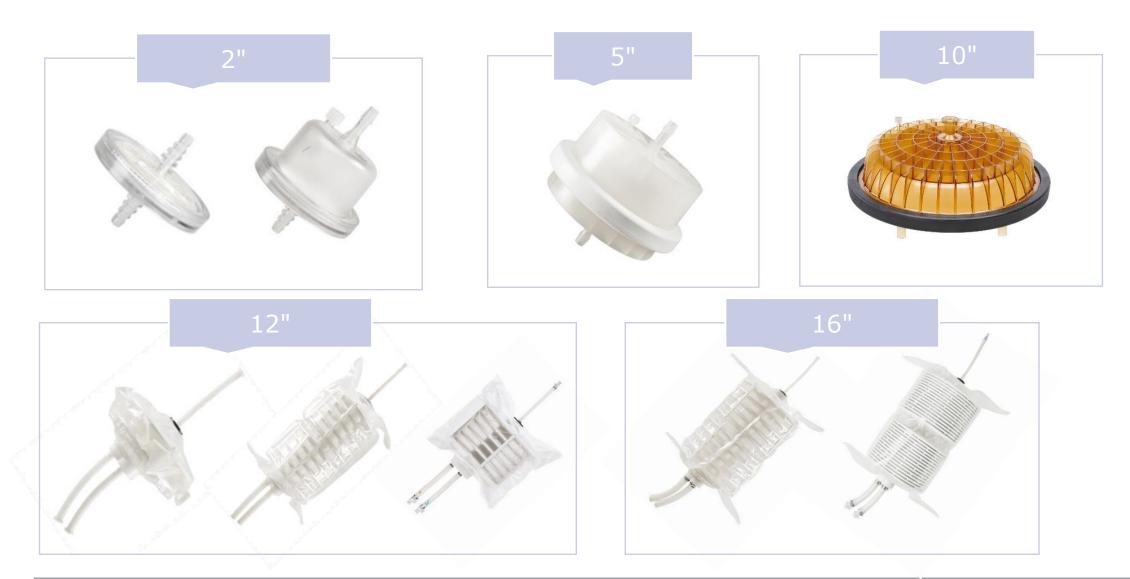


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DEPTH FILTRATION FOR VALUABLE LIQUIDS | SINCE 1938.

FILTRODISC[™] BIO SD SERIES





LABORATORY SCALE SET-UP





Some important test parameters:

- Pump need to keep the flow even with increasing pressure
- Tube connection as short as possible
- Follow the instruction in the handbook or in the playbook alluvial filtration

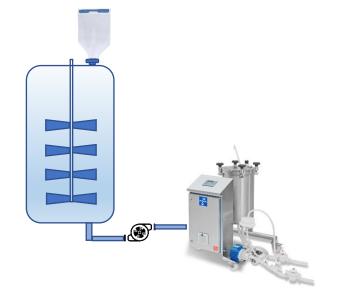
POWDER TRANSFER

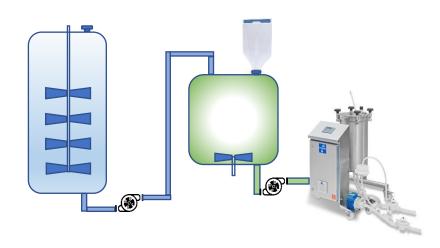


Powder Transfer into the Process

Direct mixing with fermentation broth / cell homogenate

 Feeding filter aid into bioreactor (only if single-use bioreactor) 2. Transfer fermentation broth into mixing unit and feeding filter aid



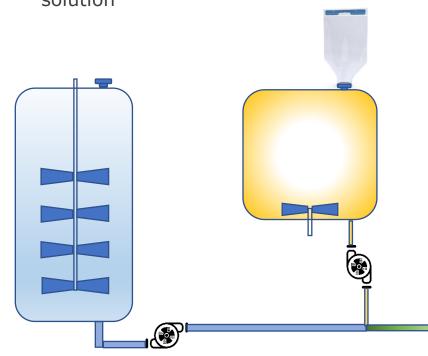


POWDER TRANSFER

Powder Transfer into the Process

Direct mixing with fermentation broth / cell homogenate

3. Body feed with a pre-prepared slurry solution



Mixing buffer with filter aid in a single-use mixer and add the solution into the line to the filter



CASE STUDY: VIRAL VECTOR CLARIFICATION



Background: Adeno-associated viruses (AAV) and lentiviruses are currently being developed for numerous indications in field of gene therapy.

 \rightarrow First step in both direct and cell-based gene therapy = packaging a therapeutic transgene into a delivery vehicle (e. g. a viral vector), followed by expansion of host cell lines to produce high-enough vector concentrations

